

RESEARCH OF REPARATIVE MECHANISMS IN THE OPTIC NERVE IN TOXIC NEUROPATHY CAUSED BY Cr (VI)

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ARTICLE INFO

Article history:

Received date 17.11.2019

Accepted date 18.12.2020

Published date 30.12.2020

Section:

Practical medicine

DOI

10.21303/2313-8416.2020.001549

KEY WORDS

chromium-induced neuropathy
oxidative stress
heat shock proteins
global pollution

ABSTRACT

Intoxication lesions of the optic nerve (toxic optic neuropathy, TON) most often occur under the influence of exogenous factors, including heavy metals. Cell survival under stress have involves heat shock proteins (HSPs).

The aim of the research. To assess the optic nerve's immunoreactivity to heat shock proteins of the HSP70 and HSP90α families and reveal its relationship with the severity of morphological changes in toxic optic neuropathy caused by Cr (VI).

Materials and methods. The study was conducted on 48 mature male rats. The experimental groups were given to drink water with Cr(VI) for 20, 40 and 60 days. This type of water is typical for the water basins in the northern districts of the Sumy region. Optic nerves changes under the influence of Cr(VI) have investigated by the morphometric method. Neuroglial cells and capillary endothelial cells were assessed by immunohistochemistry by HSP70α and HSP90 expression for intensity and spatial distribution.

Results. The data analysis revealed that Cr (VI) has a neurotoxic effect on the optic nerve with the development of edema, which is manifested by the thickening of nerve fibers. The dynamics of HSP70 immunoexpression in the endothelium of the optic nerve capillaries of rats on 20 and 40 experimental days was characterized by stable values and was 1.5 times higher than the control. The maximum number of positively stained cells for the HSP70 marker was detected in endothelial cells of the microvasculature for 60 days – 82.44±12.42 %. HSP70 levels in neuroglia cells of optic nerve have decreased on day 40 (55.66±11.56%, $p=0.05$) and lower than the control (70.44±4.81 %) group. Optic nerve capillaries was highest immunoactivity on HSP90 in group II endothelial cells – 51.22±14.57 % ($p=0.05$). The activity of HSP90α protein in optic neuroglia cells was characterized by a gradual increase in the duration of the experiment and was higher by 12.4 % in experimental group III (81.77±21.67 %) compared with control (71.66±4.95 %).

Conclusions. Our study provides an insight into the significant difference in the immunoreactivity of heat shock proteins of the HSP70 and HSP90α families in neuroglia and endothelial cells of the optic nerve capillaries under the influence of Cr(VI).

The results obtained suggest that Cr (VI) has a neurotoxic effect on the optic nerve with the development of edema, which is manifested by the thickening of nerve fibers. A comparison of the dynamics of the development of the dystrophic process in the optic nerve with the results of the immunohistochemical analysis showed, that an increase in the thickness of nerve fibers is accompanied by an increase in immunoreactive neuroglial cells (HSP90α) and endothelial cells (HSP70).

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1. Introduction

1. 1. The object of the research

Heat shock proteins (HSP70 and HSP90α) expression during experimental toxic optical neuropathy induced by Cr(VI).

1. 2. Problem description

Intoxication lesions of the optic nerve (toxic optic neuropathy, TON) most often occur under the influence of exogenous factors [1, 2], including heavy metals [3]. As the process progresses,

develop acute or chronic progressive death of retinal ganglion cells and their axons, which leads to morphological changes.

Over the past few decades, emissions of pollutants into the environment have increased significantly due to rapid industrialization, urbanization, and overuse of agricultural fertilizers [4, 5]. One of the 14 most toxic heavy metals that pollute the environment, according to the Environmental Protection Agency India, is hexavalent chromium (Cr (VI)), due to its widespread use in the industry [6]. The lack of proper control over the disposal of Cr (VI) waste leads to an increase in the level of Cr (VI) in soil, water, air, polluting the environment [6, 7]. The most common routes of exposure to chromium through these media are ingestion, inhalation, and dermal contact with soil, water, or particulates in the air contaminated with chromium.

Together with cobalt, hexavalent chromium compounds are widely used in medicine to create metal components of endoprostheses. A significant disadvantage of these prostheses is the formation of submicroscopic friction products [8, 9], toxic to the body, due to the release of heavy metal ions, particularly cobalt and chromium [10], due to implant wear and corrosion [11, 12]. A number of works carried out worldwide speak about the pathological influence of ions of cobalt and chromium on an organism [13, 14]. Of interest are ophthalmic complications, of which, to date, only isolated cases have been published in the medical literature [15, 16]. According to the Garcia M. D. and others, the Cr(VI) toxic role have been remains unclear [17].

Data from the literature do not provide a complete answer to the question of the points of application and the main mechanisms of development of disorders in the optic nerve due to the entry of hexavalent chromium into the body.

1. 3. Suggested solution to the problem

Resisting apoptosis is unthinkable without heat shock proteins (HSPs) due to their participation in a wide range of stress conditions, including pollution of the environment with heavy metals [18, 19]. The most studied of them are ATP-dependent chaperones with molecular weights from 40 to 105 kDa. These include the well-known chaperones of the 70 and 90 kDa families. The main functions of folding, refolding, elimination of irreversibly denatured protein conglomerates, or accompaniment into lysosomes (chaperone function for other proteins) to ensure homeostasis, growth processes and cell differentiation [20, 21].

In the intact retina of amphibians, the expression of HSP70 was observed mainly in the outer layers of the retina, and HSP90 – in the inner, mostly in Mueller cells and optic nerve, while ensuring the adaptive stability of its intracellular structures [22]. On immunohistochemical, HSP70 is almost not expressed in neurons, but Tytell M. and co-authors first found that this polypeptide is secreted by neuroglia cells and transported to nerve axons, thereby performing a cytoprotective function, being in adjacent cells [23].

The main aim of the work. To assess the optic nerve's immunoreactivity to heat shock proteins of the HSP70 and HSP90 α families and reveal its relationship with the severity of morphological changes in toxic optic neuropathy caused by Cr (VI).

2. Materials and methods

The rats have been kept in vivarium conditions from February to March 2018 (the vivarium of Medical Institute of Sumy State University). The study was carried out on white, unpedigreed male rats weighing 180–200 g at four months ($n=48$). The conditions of the Declaration of Helsinki (General Assembly of the World Medical Association, 2008), norms and principles of the European Convention for the Protection of Vertebrate Animals used for research and other scientific purposes were strictly observed.

2. 1. Experimental procedures

Toxic optical neuropathy in animals was simulated by adding $K_2Cr_2O_7$ to ordinary drinking water at a dose of 0.02 mol/l, which is most typical for Ukraine's contaminated regions.

For studying the dynamics of changes in the optic nerve, the experimental animals were divided into three groups, depending on the duration of taking ordinary water enriched with an increased concentration of Cr (VI). – Group I ($n=8$) – 20 days from the beginning of the experiment,

Group II ($n=8$) – 40 days and Group III ($n=8$) – 60 days, respectively. Each experimental group contained 8 control rats that drink plain water.

After the experiment's expiry, the animals were decapitated under ether anesthesia and their eyes were enucleated.

For histomorphometric examination of the optic nerve, laboratory rats were placed in a vessel where the organ was fixed in a 10 % buffered solution of neutral formalin for 24 hours. Then dehydration of the organ was performed. On a rotary microtome, serial paraffin sections of the optic nerve 5×10^{-6} m thick were obtained, which were placed on a glass slide and stained with hematoxylin and eosin.

The obtained preparations were examined, photographed and measured using a microscope "Carl Zeiss Primo Star" (Germany) (binoculars $\times 10$, lenses $\times 10$, $\times 40$) with a digital camera "Zeiss AxioCam ERC 5s" (Germany) and software package output image system and calculation of "ZEN 2 (blue edition)" (Germany).

Morphometric measurements were performed using the Digimazer program. All micro-metric indicators are presented in units of length according to the International System of Units – 10^{-6} m (μm).

For immunohistochemical study, truncated serial samples of the optic nerve were applied to glasses with high adhesiveness SuperFrost (Thermo Scientific, USA). The slides together with the material were placed in a thermostat, where they were dried at 37°C for 18 hours. The obtained samples were subjected to dehydration after dewaxing. Unmasking was performed in citrate buffer (pH 6.0) at a temperature of $95\text{--}98^\circ\text{C}$. The UltraVision Quanto Detection System HRP Polymer (Thermo scientific, USA) was used to visualize the primary antibodies. Blocking of endogenous peroxidase was performed with 3 % hydrogen peroxide solution. As a chromogen used diaminobenzidine (Thermo Scientific, USA). After the immunohistochemical reaction, sections were stained with Mayer's hematoxylin for better differentiation of tissue structures. Branches of the optic nerve were processed using polyclonal antibodies in a 1:200 dilution to HSP90 α and Hsp70 protein (Thermo Scientific and Abcam, USA).

Evaluation of the immunohistochemical reaction results was performed by calculating the area of expression (the ratio of the area of immunopositive cells to the total area of all cells expressed as a percentage) of cell elements. The reaction was considered positive in the presence of positively stained cells more than 1 % in 10 fields of view at a magnification of the microscope $\times 400$.

The images were captured by a computer-assisted digital camera, "Zeiss AxioCam ERc 5s", connected to the microscope.

All statistical analyses were performed using the Statistical Package for Social Science program (SPSS for Windows, version 15.0, SPSS Inc., Chicago, IL, USA). Determining the reliability of differences was performed using Student's t -test (t). The value of $P < 0.05$ was considered as significant.

3. Results

The results of the immunohistochemical examination of optic nerve samples of experimental rats of all groups showed different expression of HSP70 (**Fig. 1**) and HSP90 α (**Fig. 2**) in neuroglia cells and capillary endothelial cells depending on the duration of the experiment (**Table 1**).

Table 1

The content of HSP70 and HSP90 proteins in neuroglia and endothelial cells of the optic nerve capillaries of experimental animals under the influence of hexavalent chromium (Cr (VI)), %, ($M \pm m$; $n=8$)

Animal groups	Type of cells	HSP70	HSP90 α	The size of the nerve fiber, μm
Control	Endothelial cells	16.11 \pm 12.93	15.55 \pm 11.57	10.4 \pm 2.1
	Neuroglia	70.44 \pm 4.81	71.66 \pm 4.95	
Group I (20 days)	Endothelial cells	25.33 \pm 12.49	12.77 \pm 8.83	11.3 \pm 3.2
	Neuroglia	65.88 \pm 10.36	76.33 \pm 10.80	
Group II (40 days)	Endothelial cells	24.44 \pm 11.57	51.22 \pm 14.57*	13.4 \pm 2.1
	Neuroglia	55.66 \pm 11.56*	79.11 \pm 20.41	
Group III (60 days)	Endothelial cells	82.44 \pm 12.42	18.33 \pm 12.93	15.7 \pm 1.3
	Neuroglia	79.66 \pm 17.22	81.77 \pm 21.67	

Note: * – statistical significance was set at $p=0.05$

The control group of rats had a small number of HSP70 and HSP90 α positive endothelial cells in the capillaries; their number was statistically insignificant. Simultaneously, in the optic neuroglia cells observed a high level of positive cells HSP70, HSP90 α at $p<0.05$.

The dynamics of HSP70 immunoexpression in the endothelium of the optic nerve capillaries of rats of I and II experimental groups was characterized by stable values and was 1.5 times higher than the control. The maximum number of positively stained cells for the HSP70 marker was detected in endothelial cells of the microvasculature for 60 days ($82.44\pm12.42\%$), which was regarded as a strongly positive reaction. Optic nerve neuroglia cells of all study groups expressed HSP70 in almost the same amount according to the control, except for the experimental group II, where this protein's activity was equal to $55.66\pm11.56\%$ and was lower than the control $70.44\pm4.81\%$.

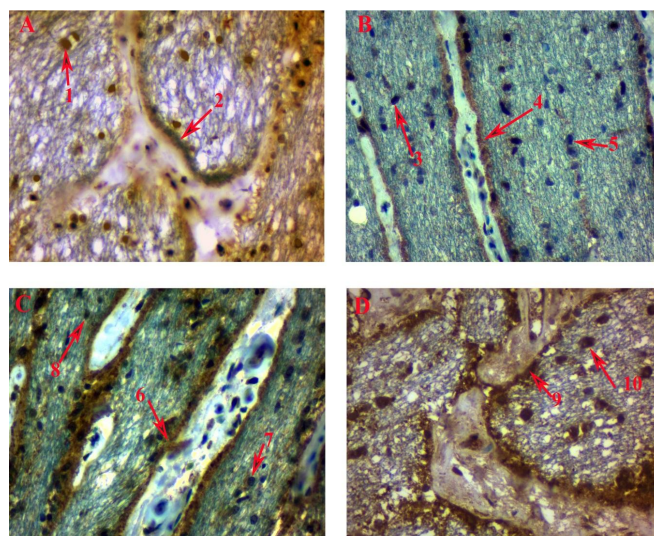


Fig. 1. Longitudinal section of the intracranial optic nerve of rats exposed to hexavalent chromium (Cr (VI)) at different time intervals. Immunohistochemical study of HSP70 expression: magnification $\times 400$: A – Control group: 1 – astrocytes; 2 – endothelium of the capillaries of the optic nerve; B – I experimental group (20 days): 3 – HSP70 – negative astrocytes; 4 – capillary endothelium with moderate expression of HSP70; 5 – HSP70 – positive cytoplasmic reaction of astrocytes; C – II experimental group (40 days): 6 – capillary endothelium with moderate expression of HSP70; 7 – HSP70 – positive cytoplasmic reaction of astrocytes; 8 – HSP70 negative astrocytes; D – III experimental group (60 days): 9 – capillary endothelium with moderate expression of HSP70; 10 – HSP70 positive cytoplasmic reaction of astrocytes

Regarding the immunohistochemical activity of HSP90 α in the endothelial cells of the optic nerve capillaries, it was highest in the II experimental group and was equal to $51.22\pm14.57\%$, followed by a sharp decrease to $18.33\pm12.93\%$ in the III group of the study and almost approaching it to the level of control $15.55\pm11.57\%$ at $p<0.05$. The activity of HSP90 α protein in optic neuroglia cells was characterized by a gradual increase in the duration of the experiment and was higher by 12, 4 % in experimental group III ($81.77\pm21.67\%$) compared with control ($71.66\pm4.95\%$).

The results of morphometric analysis indicate the presence of edema of the optic nerve, which is reflected in an increase in the thickness of nerve fibers (Table 1). In some parts of the optic nerve revealed the phenomena of focal stratification with the phenomena of precellular edema.

As a result of the correlation analysis, we found an average positive relationship between the thickness of the optic nerve fiber and the expression of HSP70 in capillary endothelial cells – $r=0.88$, at $p=0.01$. Also, a strong positive correlation was found between HSP90 α in neuroglial cells and the thickness of the nerve fiber – $r=0.94$, at $p=0.05$.

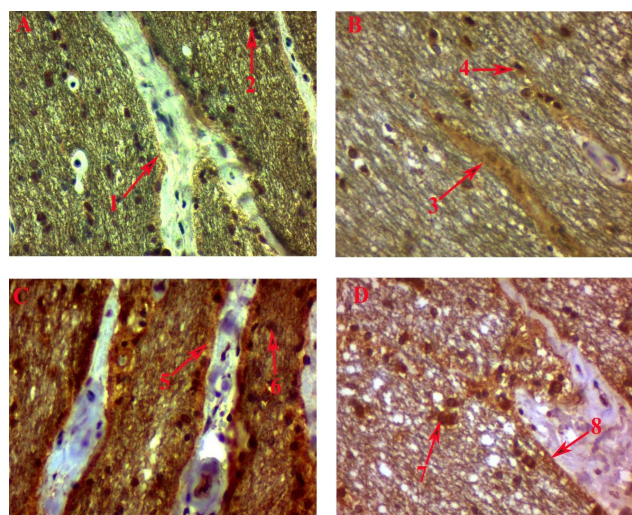


Fig. 2. Longitudinal section of the intracranial optic nerve of rats exposed to hexavalent chromium (Cr (VI)) at different time intervals. Immunohistochemical study of HSP90 α expression: magnification $\times 400$. A – Control group: 1 – capillary endothelium with weak expression of HSP90 α ; 2 – HSP90 α – positive cytoplasmic reaction of astrocytes; B – I experimental group (20 days): 3 – endothelial reaction of HSP90 α ; 4 – positive cytoplasmic and nuclear reaction of HSP90 α of astrocytes; C – II experimental group (40 days): 5 – significant endothelial expression of HSP90 α in capillaries; 6 – positive cytoplasmic, and nuclear reaction of HSP90 α astrocytes; D – III experimental group (60 days): 7 – endothelial expression of HSP90 α with dyscirculatory changes; 8 – positive cytoplasmic and nuclear reaction of HSP90 α astrocytes against edematous changes

4. Discussion

For the first time, our study describes the features of the immunoexpression of heat shock proteins of the families: HSP70 and HSP90 α in optic neuropathy associated with Cr (VI).

Some researchers believe that oxidative stress with the formation of ROS and activation of apoptosis resulting from mitochondrial dysfunction is a key link in the degeneration of retinal ganglion cells and their axons in the optic nerve [24, 25]. According to the literature, Cr (VI) enhances the accumulation of ROS by blocking complex I of the mitochondrial respiratory chain [26], and also reduces the activity of antioxidant enzymes [27]. Cr (VI), like other cationic metals, penetrates the outer mitochondrial membrane through the mechanism of molecular mimicry [28, 29]. Due to the high content of lipids with polyunsaturated fatty acids, brain mitochondria membranes are more sensitive to free radical oxidation under pathological conditions [30].

Heat shock proteins HSP70/HSP90 resist apoptosis, which in the early stages are able to reduce the consequences of free radical damage to mitochondrial membranes. It is HSP70, and according to other data, HSP90 [31], binds to Apaf-1, blocking the formation of the Apaf-1 – cytochrome C complex in the cytoplasm and disrupting the Apaf-1 bond with procaspase-9, that is, disrupting the very process of apoptosome formation [32]. It should be recalled that ROS overproduction promotes activation of the internal molecular pathway of cell apoptosis, which is regulated in mitochondria [33]. Activation of the internal pathway of apoptosis leads to an increase in the permeability of the mitochondrial membrane due to the opening of the pores and the release of cytochrome C into the cytoplasm, which binds to the activating factor of apoptotic protease – (Apaf-1), which leads to the formation of an apoptosome. Which, in turn, mediates conformational changes and activation of the initiator caspase – 9. As a result, activated caspase – 9 triggers the caspase cascade of programmed cell death via procaspases 3 and 7 [34].

In our study, we observed a decrease in the chaperone activity of HSP70 in optic neuroglia cells already in experimental group I. These results can be partially explained by previous studies, which claim that Cr (VI) activates caspase-3, thereby inducing ROS-dependent decreases in HSP70 and HSP90 [26]. However, as can be seen from our experimental results, the expression of HSP90 in glial cells, on the contrary, moderately increased and reached a peak in experimental group III,

which only partially coincides with previous studies. A simple interpretation of this is that overexpression of heat shock proteins is part of the cellular stress response and is mainly caused by the heat shock factor (HSF) [35]. Under non-stress conditions, HSF-1 is found in the cytoplasm as an inactive monomer in a complex with HSP90. When a cell is exposed to stress, HSF-1 trimerization occurs, followed by its entry into the nucleus, where it interacts with transcription elements and triggers the process of HSP 90 gene expression [36]. At the same time, the level of other heat shock proteins, including HSP70, remains low. This fact we can explain as the competition of HSPs for binding to HSF-1. In support of this, Fei Dou and coauthors [37] found increased expression of Hsp70 in the primary culture of neurons in the rat embryonic brain (E17), upon administration of an inhibitor of HSP 90, explaining this by the release of heat shock factor 1 (HSF1) from the complex with Hsp90 [38]. Termination of the harmful stimulus leads to rapid deactivation of HSF-1 and return to an inactive form. And in general, the described changes confirm the mechanism of autoregulation of heat shock protein synthesis according to the feedback principle [39].

In addition, the protective mechanism of HSP90 α is also associated with the activation of cellular signalling pathways for key proteins for which it is a chaperone [40]. Induction of HSP90 α has been reported to increase cell survival under oxidative stress caused by pollutants, incl. and heavy metals by activating signalling pathways such as JAK-STAT, ERK 1/2, PI3K/Akt, Bcl-2 and NF- κ B [41]. Activated MAP kinase, associated with the trk downstream signalling cascade, was found to coimmunoprecipitated with optic nerve HSP90, suggesting that HSP90 may be utilized in retrograde transport of the secondary messengers associated with neurotrophin signalling. HSP90 can thus be hypothesized to play a role in bidirectional RGC axonal protein transport [42].

We found an opposite picture in the capillary endothelium. Namely, overexpression of HSP70 with prolongation of the experiment time, which, as we hypothesize, is caused to maintain the integrity of the blood-brain barrier under stressful conditions. Namely, overexpression of HSP70 with prolongation of the experiment time, which, as we hypothesize, is caused to maintain the integrity of the blood-brain barrier under stressful conditions. We came to this conclusion based on its functions. In addition to blocking the caspase-dependent apoptosis pathway mentioned above, Hsp70 can directly bind the apoptosis-inducing factor (AIF), thereby preventing the caspase-independent apoptosis pathway [43]. The mechanism for triggering apoptosis by AIF is the activation of an endonuclease that cleaves nuclear DNA [44]. Kondrikov D. and others have shown that increased expression of Hsp70 under conditions of hyperoxia protects endothelial cells by inhibiting the AIF-dependent pathway of apoptosis [45]. The current result of our study is consistent with the conclusion of R. R. Shivers and co-authors [46], who demonstrated the in vitro induction of HSP70, HSP90 and HSP100 in endothelial cells of the brain microvessels in response to heat shock. They also notice – inhibition of further tight junction assembly and the disappearance and/or disassembly of tight junctions in primary cultures of bovine brain microvessel endothelial cells. It should be noted that the endothelial cells of the microvessels of the brain, together with astrocytes and pericytes, form both the blood-brain and internal blood-retinal barriers, and a violation of the relationship between them can lead to an increase in the permeability of these barriers for macromolecules and fluid from the blood to the retina and optic nerve [47].

More recently, it has been reported that mitochondria play a key role in maintaining the integrity of the blood-brain barrier (BBB) in vitro. Immunocytochemical analysis revealed that the normally well-defined, linear cell-cell junctions were disrupted when oxidative phosphorylation was inhibited by mitochondrial inhibitors [48]. It has long been suggested that mitochondrial dysfunction is associated with developing neurodegenerative diseases [49, 50].

Study limitation. Our study were limited by study of optic nerve heat shock proteins expression (HSP70, HSP90 α) under the influence Cr(VI).

Prospects for further research. In the future, it is necessary to experimentally investigate and determine changes in the retina of the eye under the influence of Cr (VI), taking into account the fact that the optic nerve is formed by axons of cells whose bodies are located in the ganglion layer of the retina.

5. Conclusions

1. Our study provides an insight into the significant difference in the immunoreactivity of heat shock proteins of the HSP70 and HSP90 α families in neuroglia and endothelial cells of the optic nerve capillaries under the influence of Cr(VI).

2. The results obtained suggest that Cr (VI) has a neurotoxic effect on the optic nerve with the development of edema, which is manifested by the thickening of nerve fibers. A comparison of the dynamics of the development of the dystrophic process in the optic nerve with the results of the immunohistochemical analysis showed, that an increase in the thickness of nerve fibers is accompanied by an increase in immunoreactive neuroglial cells (HSP90 α) and endothelial cells (HSP70).

Conflicts of interest

The authors declare that they have no conflicts of interest.

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